

**IDENTIFICATION BY MASS SPECTROMETRY OF N- β -ASPARTYL-
AND N- γ -GLUTAMYL-2-DEOXY-2-ACETAMIDO- β -D-GLUCOSYL
AMINE DERIVATIVES (*)**

L. Mester, A. Schimpl and M. Senn

Institut de Chimie des Substances Naturelles, Gif-sur-Yvette
Centre National de la Recherche Scientifique, Paris, France.

(Received 16 February 1967)

It has been reported that the carbohydrate-peptide linkage in ovalbumin (1-4), ovomucoid (5, 6), γ -globulin (7-9), α_1 -acid glycoprotein (10-12) and fibrinogen (13, 14) is formed by N- β -aspartyl-2-deoxy-2-acetamido-D-glucosylamine. The structure of this compound has been confirmed by synthesis (15, 16). However, the occurrence of other similar linkages, formed e.g. by D-glucosamine (or D-galactosamine) and glutamine, is also possible.

The aim of the present work is to develop a general method for the detection and identification of small quantities of N-acyl-2-deoxy-2-acetamido- β -D-hexosylamine derivatives. The hexosamine-aminoacid linkage such as exists in these compounds represents a type of carbohydrate-peptide linkage which is thought to be of general occurrence in a large variety of glycoproteins.

(*) - This work was supported by a research grant (n° HE-6926) from the National Institutes of Health, Bethesda, Maryland, U.S.A.

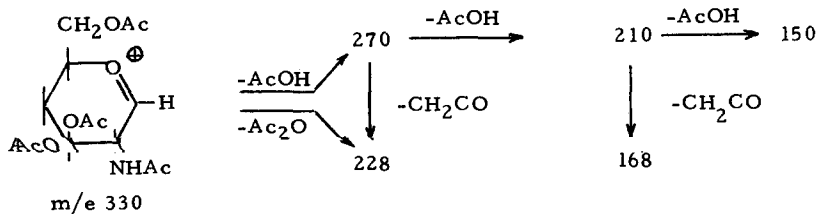
With this in view, we investigated by mass spectrometry (*) three synthetic derivatives :

(a) - N-(α -Benzyl N-benzyloxycarbonyl-L- γ -glutamyl)-2-deoxy-2-acetamido-3,4,6-tri-O-acetyl- β -D-glucosylamine (17) (I), whose structure and mass spectrum are shown in figure 1.

(b) - The corresponding L- β -aspartyl derivative (17) (II).

(c) - N-(α -Ethyl N-trifluoroacetyl-L- β -aspartyl)-2-deoxy-2-acetamido-3,4,6-tri-O-acetyl- β -D-glucosylamine (III). The structure and mass spectrum of this last compound (m.p. 252-5°), obtained by condensation of 2-deoxy-2-acetamido-3,4,6-tri-O-acetyl-D-glucosylamine with N-trifluoroacetyl-L-aspartic acid α -ethyl ester in the presence of dicyclohexylcarbodiimide, is shown in figure 2.

The amino-sugar moiety of the compounds is indicated in the spectra by a fragment at m/e 330 :



(*) - The spectra were recorded with an high resolution instrument MS9 (A.E.I.). The temperature in the ion source was about 280°C, the electron energy was set at 70 eV. The samples were introduced directly into the ion source through a vacuum lock system.

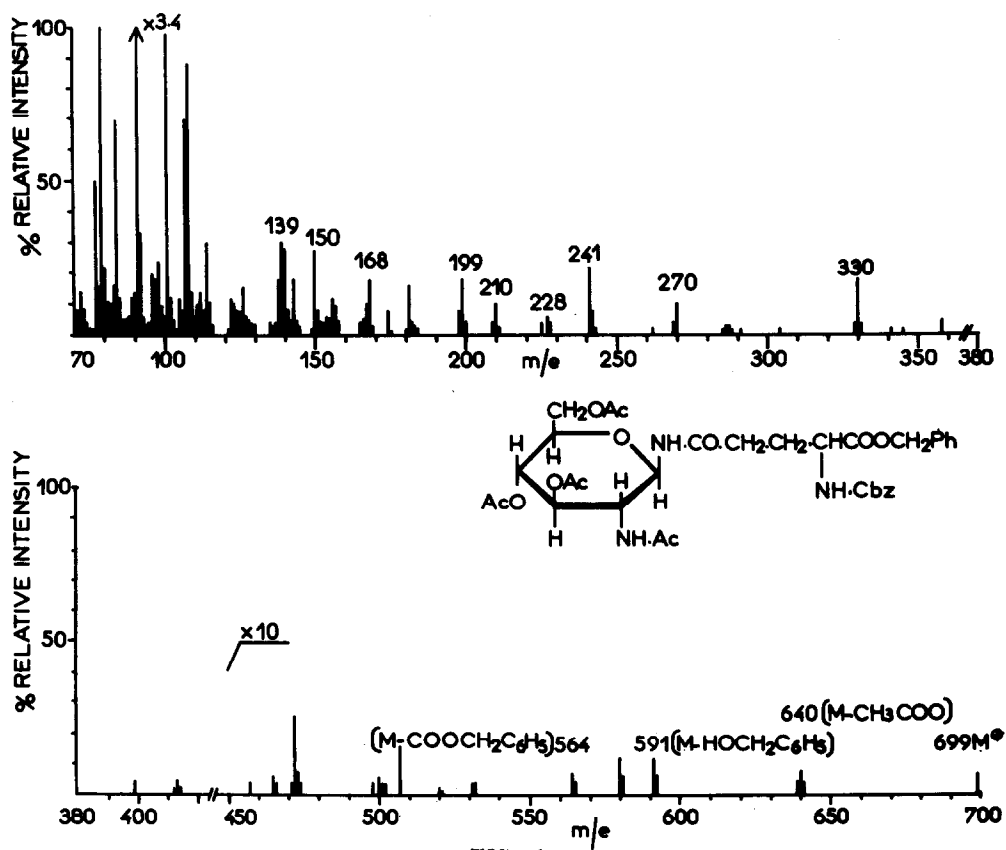


FIG. 1

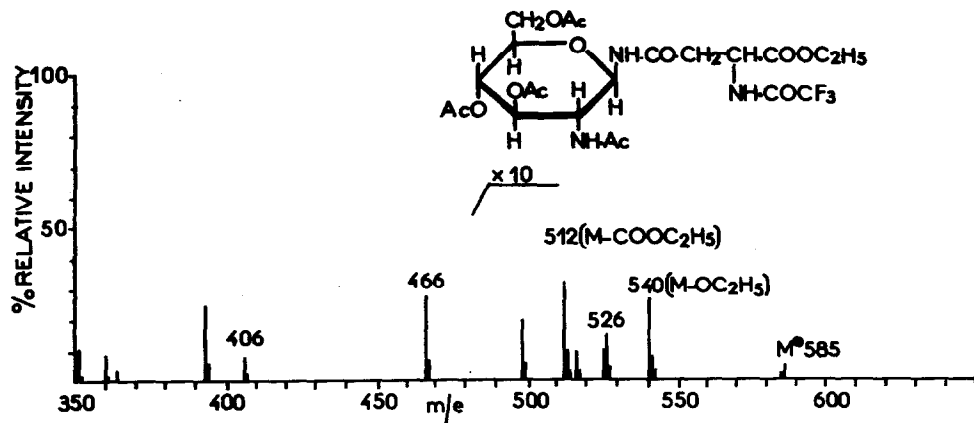
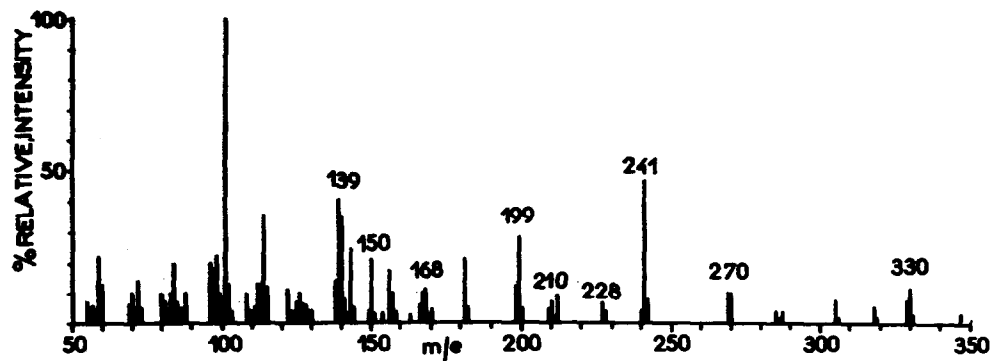
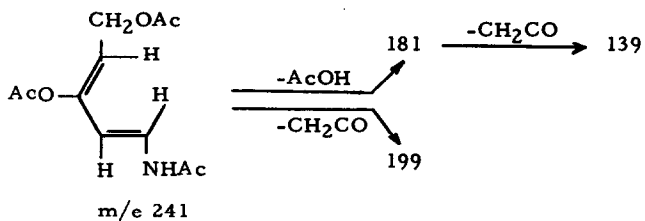


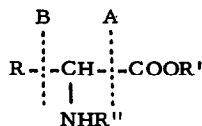
FIG. 2

Further fragmentation of the oxonium ion corresponds to the general reactions observed in peracetylated sugars (18) and peracetylated amino-sugars (19), involving the loss of acetic acid and ketene; it may be summarized by the above scheme. The fragments shown in the scheme may be accompanied by ions containing one less hydrogen atom which originate from an ion at m/e 329.

Another peak series is found starting from an ion at mass 241, which corresponds to the structure of the ion at m/e 242 in peracetylated sugars (18). The loss of acetic acid and ketene from this ion is outlined in the following scheme:



The molecular ions are found for (I) at m/e 699, for (II) at m/e 685 and for (III) at m/e 585. Characteristic peaks of most amino-acid esters are due to the loss of the ester group (20) (cleavage A):



sequence in the environment of these linkages in glycopeptides should then be feasible. The work is now being extended to natural products.

References

1. I. Yamashina and M. Makino, J. Biochem. (Tokyo), 51, 359 (1962).
2. V.P. Bogdanov, E.D. Kaverzneva and A. Andrejeva, Biochem. Biophys. Acta, 65, 168 (1962).
3. G.S. Marks, R.D. Marshall and A. Neuberger, Biochem. J., 85, 15P (1962).
4. A.P. Fletcher, R.D. Marshall and A. Neuberger, Biochem. J., 88, 37P (1963).
5. M. Tanaka, J. Pharm. Soc. Japan, 81, 1460 (1961).
6. J. Montreuil, G. Biserta and A. Chosson, Compt. Rend., 256, 3372 (1963).
7. J.A. Rothfus, Fed. Proc., 20, 383 (1961).
8. J.A. Rothfus and E.L. Smith, J. Biol. Chem., 238, 1402 (1963).
9. N. Duquesne, M. Monsigny and J. Montreuil, Compt. Rend., 262, 36 (1966).
10. E.H. Eylar, Biochem. Biophys. Res. Commun., 8, 195 (1962).
11. S. Kamiyama and K. Schmid, Biochem. Biophys. Acta, 58, 80, (1962).
12. R.C. Hughes and R.W. Jeanloz, Biochem., 5, 253 (1966).
13. L. Mester, E. Moczar, G. Vass and L. Szabados, in "Structure and Function of Connective and Skeletal Tissue" (Proc. NATO Adv. Study Inst., 1964), Butterworths, London 1965, p. 171.
14. L. Mester, E. Moczar, G. Vass and L. Szabados, Path. Biol., 13, 540 (1965).
15. G.S. Marks, R.D. Marshall and A. Neuberger, Biochem. J., 87, 274 (1963).
16. H. Tsukamoto, A. Yamamoto and C. Miyashita, Biochem. Biophys. Res. Commun., 15, 151 (1964).
17. A. Yamamoto, C. Miyashita and Tsukamoto, Chem. and Pharm. Bull. (Tokyo), 13, 1041 (1965).
18. K. Biemann, D. De Jongh and H.K. Schnoes, J. Amer. Chem. Soc., 85, 1763 (1963).
19. M. Senn and A.L. Burlingame, unpublished results.
20. H. Budzikiewicz, C. Djerassi and D.H. Williams, "Structure Elucidation of Natural Products by Mass Spectrometry", Vol. II, p. 183, Holden-Day, Inc. 1964.
21. R.T. Aplin, J.H. Jones and B. Liberek, Chem. Comm., 21, 794 (1966).